

## Modeling the cell cycle: New Skills in Undergraduate Biology Activities Education

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## Goal of today's 1.5hr

- Introduce you to what was done
- Introduce you to modeling the cell cycle
- Relationship to new skills:
  - Education reform looks at engaging students in: \_\_\_\_\_
  - In biology, scientists are having to learn new ways of understanding biological systems
  - New ways are quantitative, theoretical and conceptual

## The Lens

- Performance
- Co-teaching
- Cellular to biochemical view
  - Biology, math, models: conceptual, mathematical, computers

## What do I mean by performance?

- Not a measure, rather an activity
- Activity of becoming:
  - students are both who they are: “students”
  - and who they are not: “scientists/mathematicians”
- Vygotsky: zones of proximal development
  - Co-learning, group, social, activity
- Performatory
  - Understanding developed in practice
  - [All Stars Project, Inc.](#) and [East Side Institute](#)

## Classroom as performance

- **Directors:** John Jungck and Raquell Holmes
- **Performers:** Instructors and students
- **Play:** Modeling the cell cycle
- **Length:** Five weeks, small groups
- **Props:** Stella, former scripts (models) and text.
- **Direction:** create models and discover what you need.

## Activity: Performing as scientists

- **The background:** biology
- **The tools:** computer modeling
- **The challenge:** take what is known and create something new
  - New understanding
  - New model

## What we did and we will do

- Research Course
- Students perform as scientists
- Introduced 4-5week module on modeling the cell cycle
- Students worked in groups not from same year
  - on first exercise only
- Hands on exercise
- Perform as students/scientists
  - New to topic
  - Some understanding
  - Different skills
- Modified version of first modeling lab in class
- Work in groups not from same discipline

## Today's collective performance

- As a large group:
  - Introduction to biology
  - Introduction to modeling
- Smaller groups (2 or 3):
  - Create models of aspects of the cell cycle

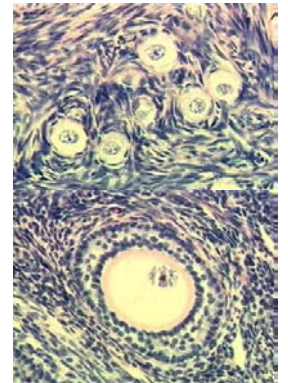
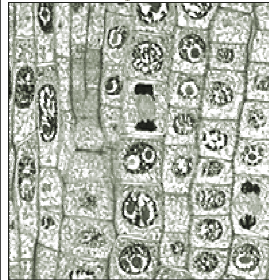
## Cell Cycle/ Cell division

- What do we know?
  - A collective version/story
- Why do we care?
  - A collective version/story

Group sharing or improvisation

## Cell: Cycles and Growth

Cells in the tip of an onion root



Gap phases (G1, G2)  
Synthesis (S phase)  
Mitosis (M phase)

## Key experiments

See Figure 1-10 of *The Cell Cycle* by Murray and Hunt, 1993.

Fusion of somatic cells in different cell cycle stages illustrates logic of cell cycle progression.

Murray and Hunt, 1993. *The Cell Cycle*

## Key experiments contd.

See Figure 2-5 of *The Cell Cycle* by Murray and Hunt, 1993.

Cytoplasmic transfer experiments demonstrates presence of maturation promoting factor: MPF.

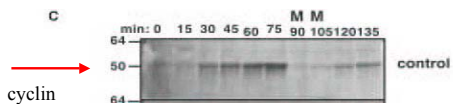
## What is MPF?

- What characteristics does it have or have to have?
  - How can this function be regulated?

## What is MPF?

- What characteristics does it have or have to have?
  - Temporally regulated function
    - “On” during M-phase, “off” during interphase
      - Biological assay
    - In embryos must cycle, show periodicity
  - How can this function be regulated?
    - Regulated synthesis
    - Regulated form (phosphorylation, protein complexes)
    - Regulated degradation

## Interesting notes



- Took a very long time to discover cyclins.
  - Discover a protein synthesized periodically
  - Discover a protein degraded periodically

Figure excerpt from Sha et. al., 2003; PNAS

## What we believe/know...

- Cyclin synthesis is constant
- MPF activity is turned “on” and “off”
- MPF activity is turned on by cyclin
- Cyclin is degraded

## Creating a Computational Model

- Concept Map
- Factors and relationships between factors
- Describe relationships mathematically
- Solve equations: using computer tools
- View and interpret results

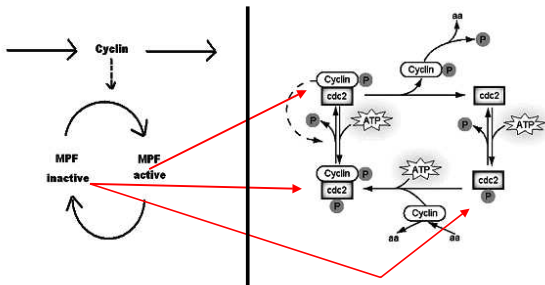
## To be performed:

See First Lab Ex.

Draw flow diagrams/concept map for the statements **provided below**. Keep your hand drawings and turn them in.

1. System statements
  - inactive MPF **becomes** active MPF
  - Active MPF **becomes** inactive MPF
2. System statements
  - Cyclin is **synthesized and degraded**
  - Cyclin **stimulates** inactive MPF **to become** active MPF

# Current conceptual models of the cell cycle

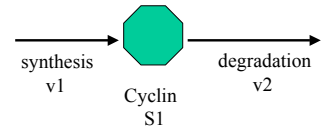


Or: Write sentences for the conceptual models

# Creating a Computational Model

- Describing relationship **mathematically**

## Representations



### Difference equations

Cyclin= synthesis -degradation

### Ordinary differential equation

$$\frac{dS_1}{dt} = v_1 - v_2$$

# Designing a dynamic experiment

- Concept Map
- Factors and relationships between factors
- Describing relationship mathematically
- What **rate laws** are known to describe the enzymatic reaction?
  - Types of rate laws/kinetic models
    - Constant, mass action, michaelis menten...
  - Initial conditions/values
    - Often useless in modeling papers
    - Opportunity to work with research papers

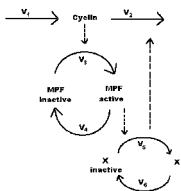
# The model we're playing with

Asks: are the minimal components of the system sufficient to account for the oscillation patterns in early embryos? Golbeter, 1991

STELLA model:

Continuous, non-stochastic, Non-spatial, population

# Base model in an overview



### Difference Equations

Cyclin= synthesis-degradation = V1-V2

MPF= activation-inactivation = V3-V4

Inactive MPF= inactivation-activation = V4-V5

X=activation-inactivation = V5-V6

### Ordinary Differential Equations

$$\frac{dC}{dt} = V_1 - V_2$$

$$\frac{dM}{dt} = V_3 - V_4$$

$$\frac{dX}{dt} = V_5 - V_6$$

### Mass Action Rate Equations

V1=constant V4=k\*MPF

V2=k\*Cyclin\*X V5=k\*MPF\*X

V3=k\*iMPF\*Cyclin V6=k\*X

Initial conditions in "Cell Cycle" chapter

# Ex contd.

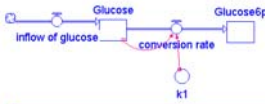
B. Create your concept maps in Stella.

C. Assume the following values for reactions and variables and enter them to your Stella model:

- All **reactions** are linear and based on the law of mass action (rate constant x substrate).
- Vary your **rate constants** and **amounts**

# Stella

Concept Map



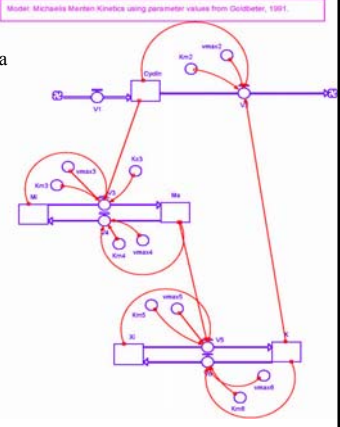
Rules as math

```
[ RUNTIME EQUATIONS ]
Glucose(t) = Glucose(t - dt) + (inflow_of_glucose - conversion_rate) * dt
Glucose6p(t) = Glucose6p(t - dt) + (conversion_rate) * dt
conversion_rate = k1*Glucose
```

Initial Conditions

```
[ INITIALIZATION EQUATIONS ]
k1 = 50(1/min)
INIT Glucose = 0
inflow_of_glucose = 50(mM*1/min)
conversion_rate = k1*Glucose
INIT Glucose6p = 0
```

A version of Goldbeter in Stella



Reference model for students

Models of cell cycle taking into account the following:

1. Cell size in yeast
2. MPF self-activation of MPF in mammalian or amphibian cells
3. Binding rates of cyclin to cdc2 in yeast, mammalian or amphibian cells
4. Phosphorylation rates of cdc2/MPF in yeast, mammalian or amphibian cells
5. Mechanisms of threshold generation: Michaelis-menten models
6. Ubiquitination and cyclin degradation mechanisms yeast, mammalian or amphibian cells
7. Additional regulators of activation or inactivation of MPF yeast, mammalian or amphibian cells

## Student models

Wee1 and Cdc25 regulation of Cell Cycle

Eq. 1  
 $2.3 \log [S]_0/[S] = kt$

S=Substrate  
 k=Rate Constant  
 t=Time

Ex. Wee1 activation constant  
 $[S]_0 = 100$   $[S] = 50$   $t = 7.5$   
 $2.3 \log (100/50) = 7.5k$   
 $k = 0.092 \text{ nM}^{-1} \text{ min}^{-1}$

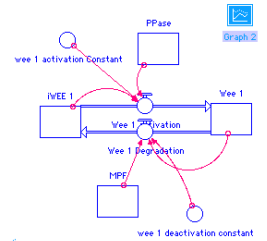
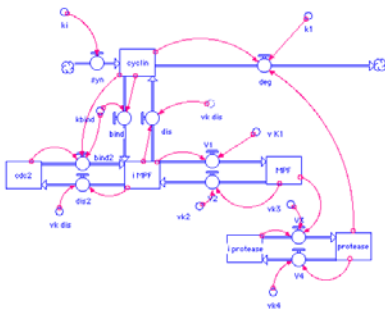


Figure 2. Wee1 model

Chung, Morgan-Wesiburg and Murphy

## Student models

We believe that our results support our hypothesis that the **cyclin-cdc2 binding** rate affects the cell cycle. As binding rate increases in relation to dissociation rate, oscillation frequency and amplitude increases; the reverse is true when dissociation rate is greater.



## Items we hit

- What does activate, bind, turn on mean in terms of a mathematical equation (+, x, /)
- How do we construct concepts in Stella?
- My results don't look like...
- Wanted more practice with equations and stella, so that they could trust their setup decisions.
- Rarely related results to rate equations